

**Supplemental Information**  
 Supplemental Tables 1 and 2  
 Supplemental Figures 1-4

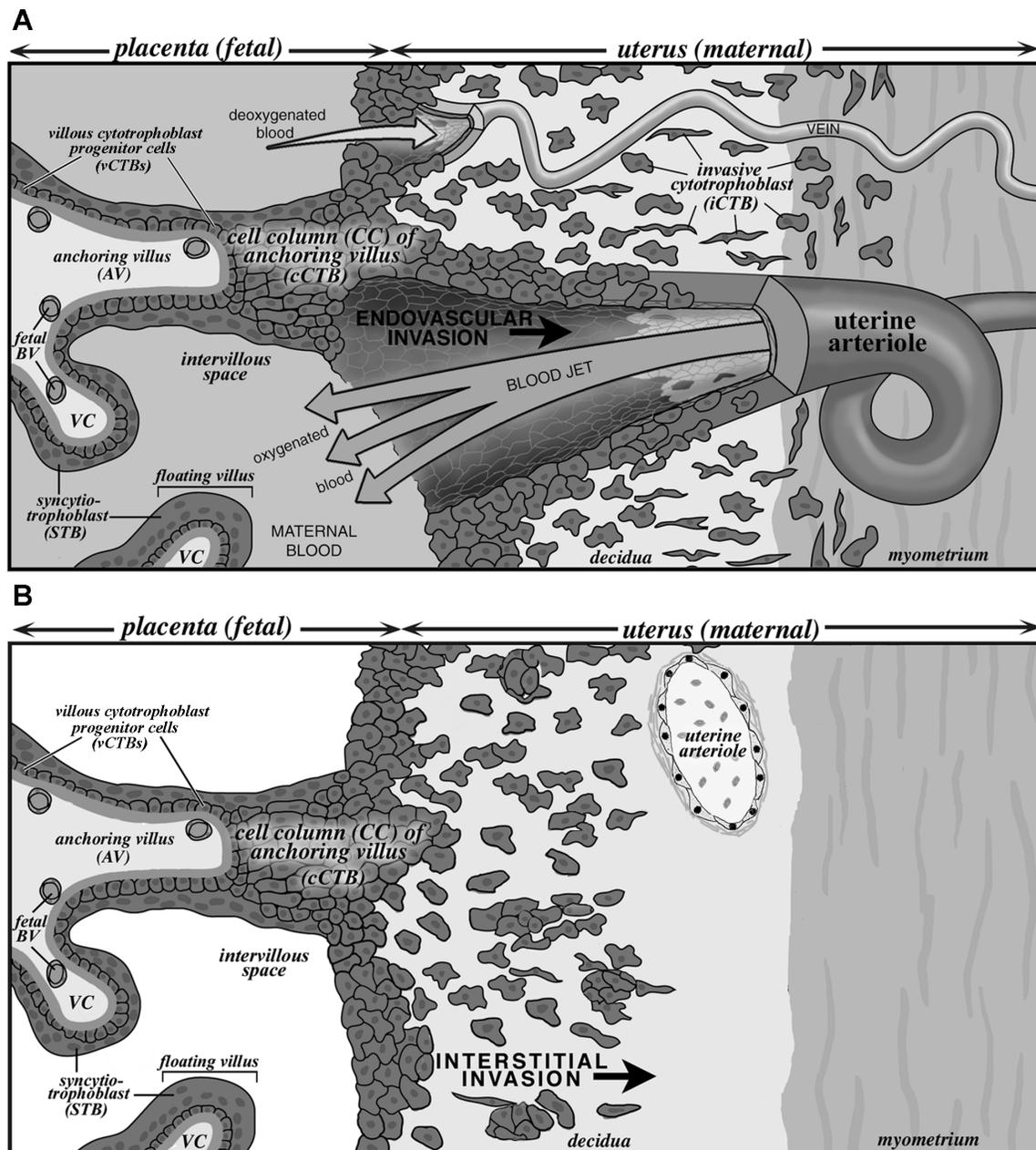
**Supplemental Table 1. Maternal and infant characteristics**

	PE ( <i>n</i> = 5)	nPTL ( <i>n</i> = 5)	<i>P</i> value
Maternal age, yr	29.0 (5.9)*	25.6 (6.3)	0.40
BMI, kg/m <sup>2</sup>	27.4 (4.2)	26.0 (5.0)	0.68
Systolic blood pressure, mmHg	148 (11)	112 (7)	< 0.001
Diastolic blood pressure, mmHg	88 (5)	65 (13)	< 0.01
Proteinuria, designation	+1 to +3	0	NA
Gestational age at delivery, week	31.2 (2.5)	29.5 (3.7)	0.45
Birth weight, g	1365 (528)	1572 (694)	0.61

\*mean ± SD, 2-tailed Student's t-test.

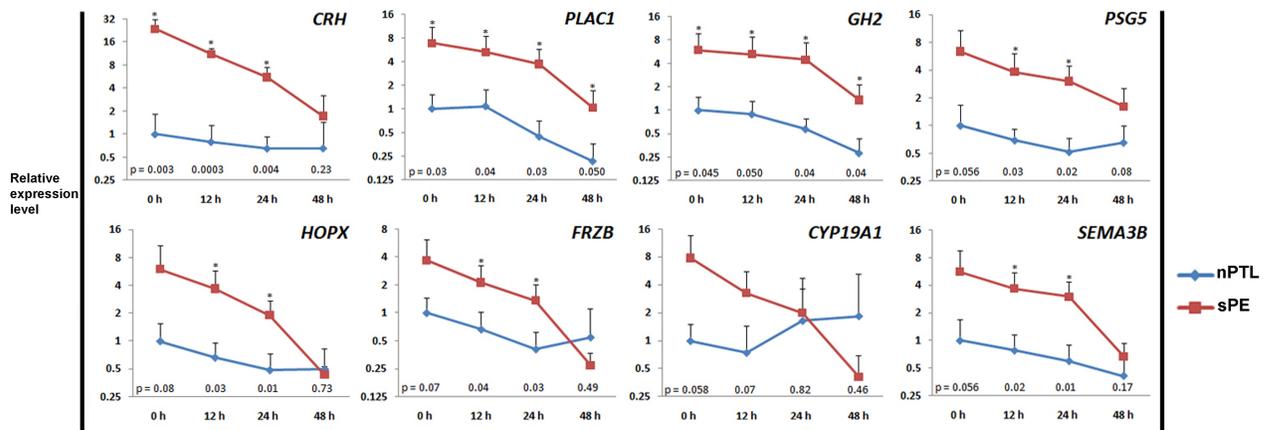
**Supplemental Table 2. Reagents, sources and concentrations**

<b>Reagent</b>	<b>Source</b>	<b>Concentration</b>
Polyclonal anti-SEMA3B	Novus Biologicals	Diluted 1:1000
Polyclonal anti-NRP-1 (C-19) and NRP-2 (C-9)	Santa Cruz Biotechnology Inc. (Santa Cruz, CA)	2 µg/ml (IB)
Monoclonal anti-VEGF R2	Dr. K. Chwalisz (Scherring AG, Berlin, Germany)	1:500
Polyclonal anti-PI3K p85	Upstate USA Inc (Charlottesville, VA)	1:100
Polyclonal anti-PI3K p100 $\alpha$	Epitomics (Burlingame, CA), Becton Dickinson (San Jose, CA)	1:2000 250 ng/ml
Polyclonal anti-Akt, anti-p-Akt (Ser473) and anti-p-GSK3 $\alpha/\beta$ (Ser21/Ser9)	Cell Signaling Technology (Danvers, MA)	1:1000 1:1000 1:1000
Monoclonal anti-GSK3 (4G-IE)	Upstate Cell Signaling Solutions (Billerica, MA)	1 µg/ml
Monoclonal anti-V5	Invitrogen Corp (Carlsbad, CA)	160 ng/ml
Monoclonal anti- $\alpha$ -actin (AC-40)	Sigma-Aldrich Corp (Saint Louis, MO)	1:500
(rat) monoclonal anti-cytokeratin (7D3)	Generated in collaboration with Dr. C. Damsky (Damsky <i>et al.</i> , 1992)	1:200
NRP1-Fc, NRP2-Fc and CD6-Fc fusion proteins	R&D Systems (Minneapolis, MN)	400 ng/ml
Monoclonal anti-VEGF (A4.6.1)	Dr. Napoleone Ferrara (Genentech, South San Francisco, CA)	400 ng/ml
Wortmannin	Calbiochem Signal Transduction (San Diego, CA)	2 µM
Complete EDTA-free Protease inhibitor cocktail	Roche Applied Science (Indianapolis, IN)	–
LiCl	Sigma-Alrich Corp	20 mM
SEMA3B	Produced in house	50-100 ng/ml (COS-1 cells), 2 µg/ml ( <i>E. coli</i> )
VEGF <sub>165</sub>	R&D Systems	40 ng/ml medium; 60 pg/filter



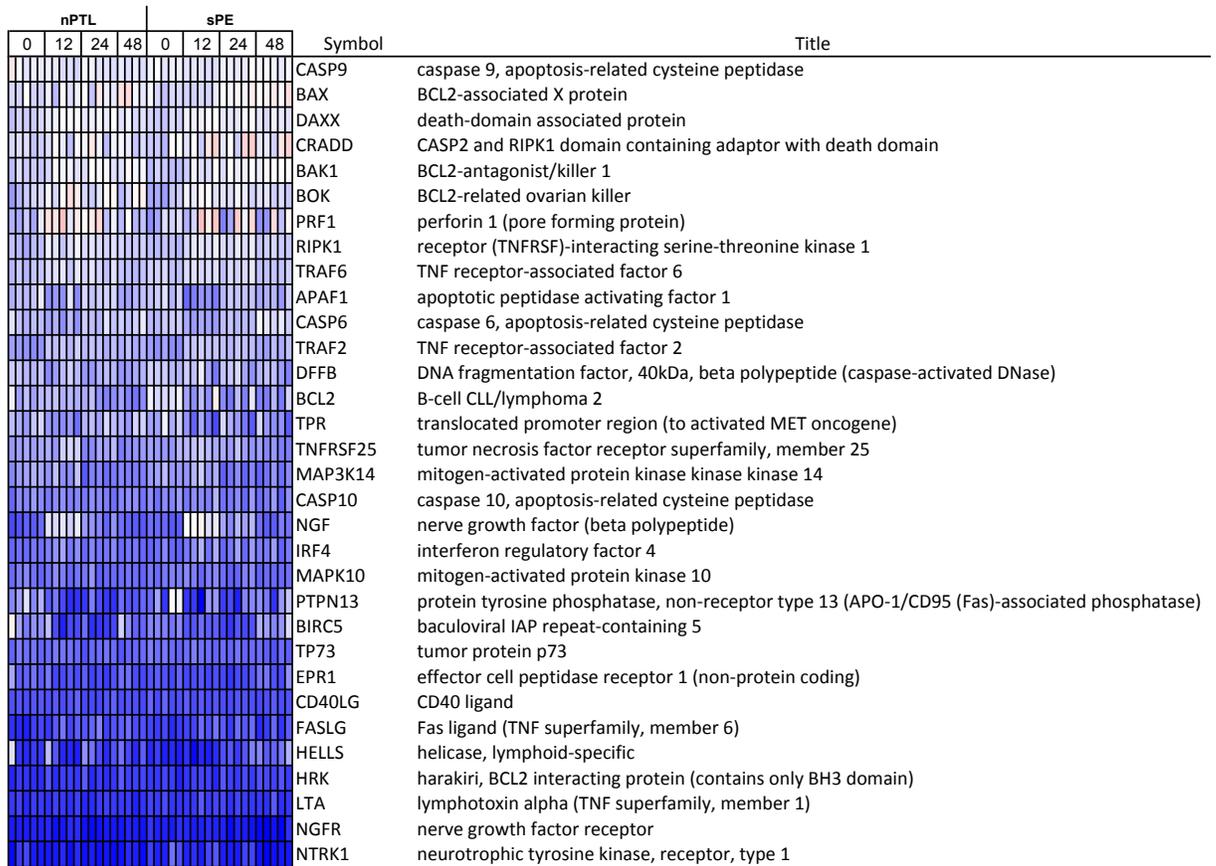
**Supplemental Figure 1.** Diagram of the cellular organization of the human maternal-fetal interface in normal pregnancy and in preeclampsia. (A) Villous cytotrophoblasts (vCTBs) progenitors, the specialized (fetal) epithelial cells of the placenta, differentiate and invade the uterine wall (interstitial invasion; iCTBs), where they also breach maternal blood vessels (endovascular invasion). The basic structural units of the placenta are the chorionic villi, composed of a stromal villous core (VC) with fetal blood vessels, surrounded by a basement membrane and overlain by vCTBs. During differentiation, these cells detach from the basement membrane and adopt one of two fates. They either fuse to form the multinuclear syncytiotrophoblasts (STBs) that cover floating villi or join a column of cytotrophoblasts (cCTBs) at the tips of anchoring villi (AV). The syncytial covering of floating villi mediates the nutrient, gas and waste exchange between fetal and maternal blood. The anchoring villi, through the attachment of cCTBs, establish physical connections between the fetus and the mother. iCTBs penetrate the uterine wall through the first third of the myometrium. A subset of these cells home to uterine spiral arterioles and remodel these vessels by replacing the

endothelial lining and intercalating within the muscular walls. To a lesser extent, they also remodel uterine veins. (B) In PE, the interstitial and the endovascular components of CTB invasion are restricted. As a result, interstitial invasion is shallow and many uterine arterioles retain their original structures.

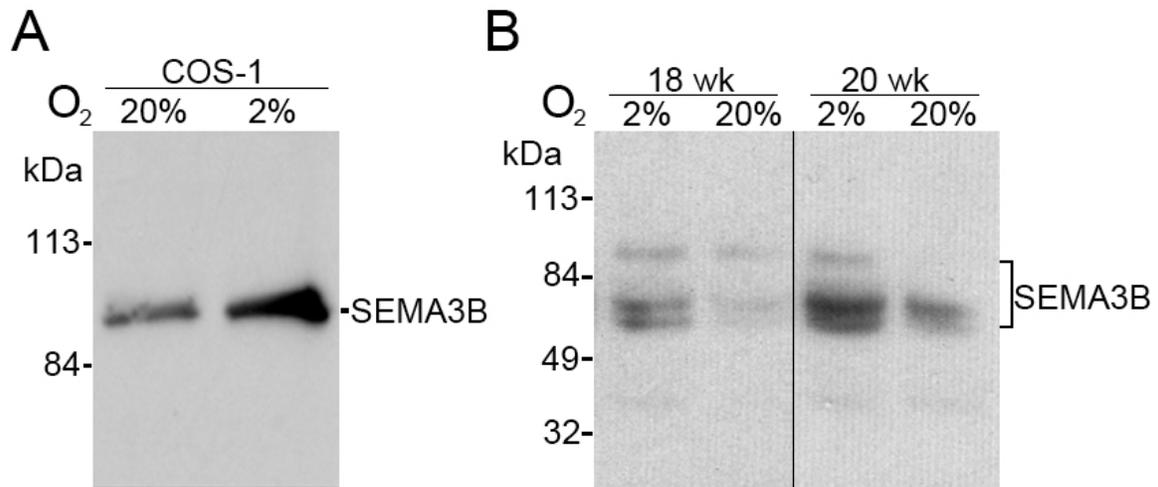


**Supplemental Figure 2. Confirmation of the expression patterns of selected genes that were misexpressed in sPE.** Taqman qRT-PCR was performed using cDNA samples prepared from CTBs that were isolated from the placentas of women whose pregnancies were complicated by preterm labor with no sign of infection (nPTL) or severe preeclampsia (sPE). The analyses were done on cells immediately after isolation (0 h) and after 12, 24, or 48 h of culture. The reactions were done in triplicate. The relative gene expression levels of each transcript were plotted on the y-axis. Data points are the mean  $\pm$  standard deviation. Fold change differences between the sPE and nPTL samples at each time point were assessed using a Student's t-test. The values shown are the mean  $\pm$  SD. p values are displayed at bottom of each panel.

nPTL				sPE				Symbol	Title
0	12	24	48	0	12	24	48		
								TNFSF10	tumor necrosis factor (ligand) superfamily, member 10
								NFKBIA	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha
								CYCS	cytochrome c, somatic
								MCL1	myeloid cell leukemia sequence 1 (BCL2-related)
								TFG	TRK-fused gene
								JUN	jun oncogene
								BIRC2	baculoviral IAP repeat-containing 2
								BCL2A1	BCL2-related protein A1
								NR3C1	nuclear receptor subfamily 3, group C, member 1 (glucocorticoid receptor)
								TNFRSF1A	tumor necrosis factor receptor superfamily, member 1A
								BNIP3L	BCL2/adenovirus E1B 19kDa interacting protein 3-like
								TNFRSF1B	tumor necrosis factor receptor superfamily, member 1B
								IRF1	interferon regulatory factor 1
								SFRS2IP	splicing factor, arginine/serine-rich 2, interacting protein
								XIAP	X-linked inhibitor of apoptosis
								CASP4	caspase 4, apoptosis-related cysteine peptidase
								IRF7	interferon regulatory factor 7
								RELA	v-rel reticuloendotheliosis viral oncogene homolog A (avian)
								CASP1	caspase 1, apoptosis-related cysteine peptidase (interleukin 1, beta, convertase)
								TNFRSF10B	tumor necrosis factor receptor superfamily, member 10b
								TNFRSF21	tumor necrosis factor receptor superfamily, member 21
								BCL2L11	BCL2-like 11 (apoptosis facilitator)
								MYC	v-myc myelocytomatosis viral oncogene homolog (avian)
								CASP3	caspase 3, apoptosis-related cysteine peptidase
								CASP7	caspase 7, apoptosis-related cysteine peptidase
								IRF2	interferon regulatory factor 2
								FADD	Fas (TNFRSF6)-associated via death domain
								BID	BH3 interacting domain death agonist
								MDM2	Mdm2 p53 binding protein homolog (mouse)
								BIRC3	baculoviral IAP repeat-containing 3
								MAP3K1	mitogen-activated protein kinase kinase kinase 1
								CASP2	caspase 2, apoptosis-related cysteine peptidase
								BCL2L2	BCL2-like 2
								NFKB1	nuclear factor of kappa light polypeptide gene enhancer in B-cells 1
								TRADD	TNFRSF1A-associated via death domain
								IRF3	interferon regulatory factor 3
								CHUK	conserved helix-loop-helix ubiquitous kinase
								IRF6	interferon regulatory factor 6
								BCL2L1	BCL2-like 1
								TRAF3	TNF receptor-associated factor 3
								NFKBIE	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, epsilon
								NFKBIB	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, beta
								MAP2K4	mitogen-activated protein kinase kinase 4
								CARD16	caspase recruitment domain family, member 16
								BAD	BCL2-associated agonist of cell death
								CASP8	caspase 8, apoptosis-related cysteine peptidase
								IKBKE	inhibitor of kappa light polypeptide gene enhancer in B-cells, kinase epsilon
								GZMB	granzyme B (granzyme 2, cytotoxic T-lymphocyte-associated serine esterase 1)
								TP53	tumor protein p53
								TNF	tumor necrosis factor
								TRAF1	TNF receptor-associated factor 1
								FAS	Fas (TNF receptor superfamily, member 6)
								CD40	CD40 molecule, TNF receptor superfamily member 5
								IKBKG	inhibitor of kappa light polypeptide gene enhancer in B-cells, kinase gamma
								PARP1	poly (ADP-ribose) polymerase 1
								IKKB	inhibitor of kappa light polypeptide gene enhancer in B-cells, kinase beta
								DFFA	DNA fragmentation factor, 45kDa, alpha polypeptide
								IRF5	interferon regulatory factor 5



**Supplemental Figure 3.** CTBs isolated from the placentas of nPTL and sPE patients did not upregulate genes that are associated with apoptosis over 48 h of culture.



**Supplemental Figure 4.** Hypoxia upregulated SEMA3B expression after 48 h in culture. Transfected COS-1 cells (A) and chorionic villus explants established from normal (18 and 20 wk) second trimester placentas (B) were cultured for 48 h in standard conditions (20% O<sub>2</sub>) or in a hypoxia chamber (2% O<sub>2</sub>). Vertical lines denote noncontiguous lanes from the same gel. The entire experiment was done a total of three times. SEMA3B expression was quantified by immunoblot analysis.